

Superresolution two-photon fluorescence microscopy using photoswitchable fluorescent proteins

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the University of Szeged, Hungary,

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and supported from

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The principal goal

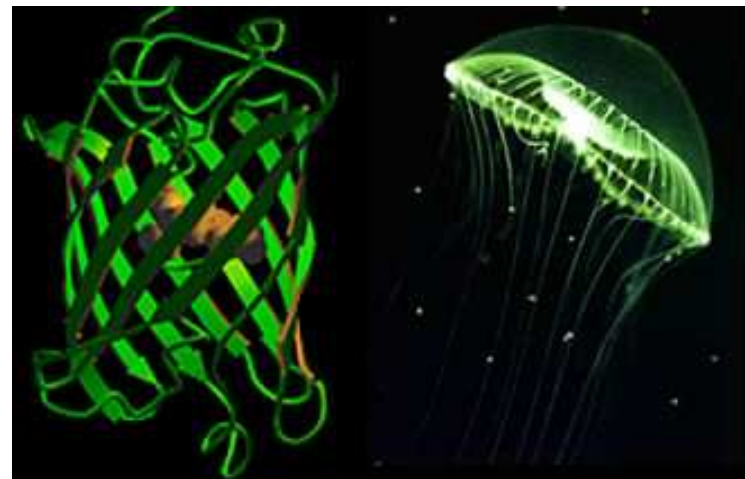
- Combine the method of two-photon polarization microscopy with microscopy techniques allow breaking the diffraction limit on resolution of optical microscopes.

Aim of the project

- Create a membrane-localized photoswitchable fluorescent protein suitable for combining two-photon polarization microscopy with STED superresolution microscopy.

What is the GFP?

- Green Fluorescent Protein
- 238 amino acids
- Fluorescence
- *Aequorea victoria*



Structure of the GFP

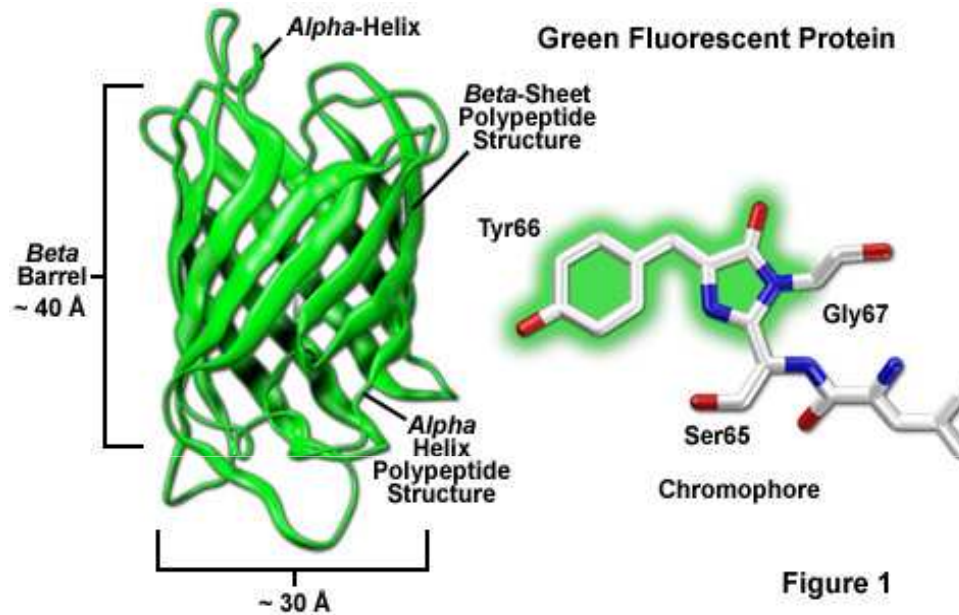


Figure 1

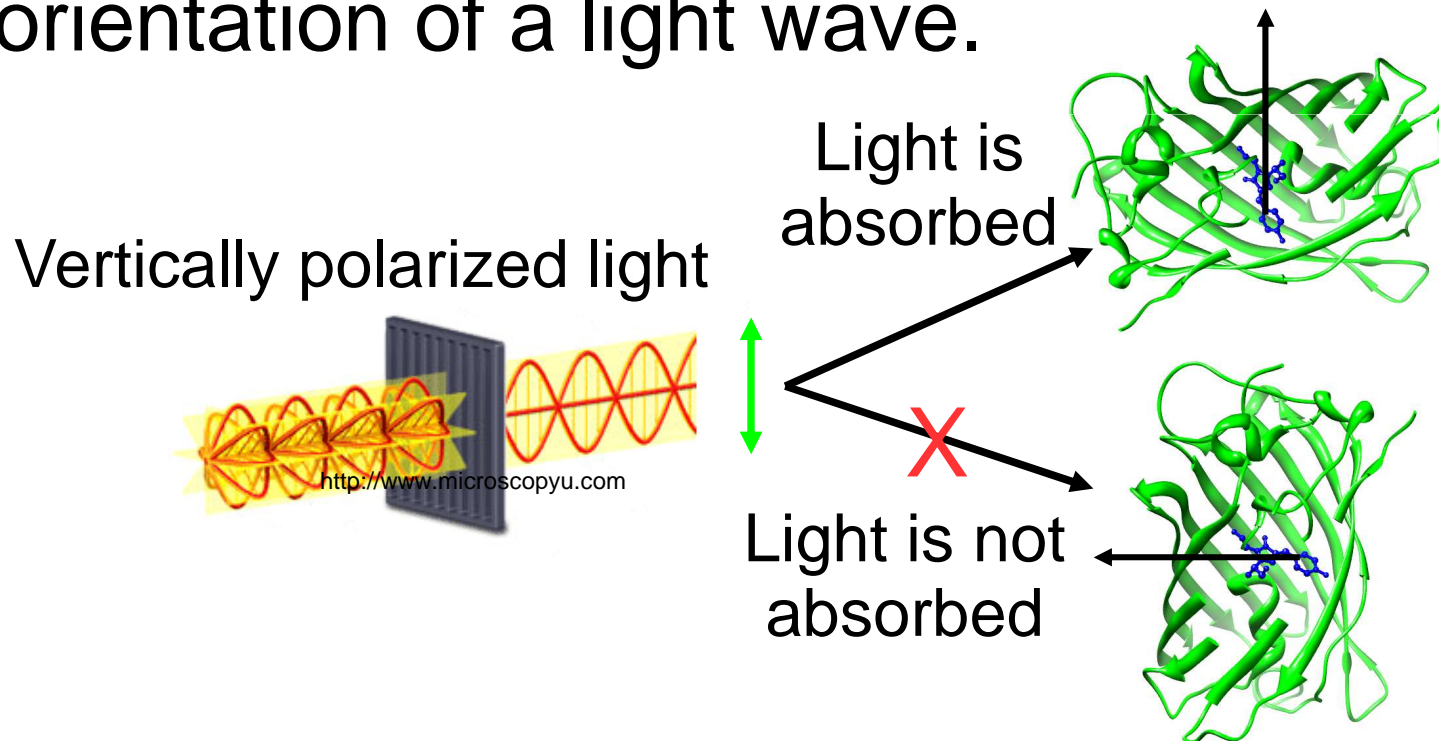
- Chromophore/fluorophore
- Planar structure

How can we use fluorescent proteins for studies of membrane proteins?



Fluorescent proteins are sensitive to excitation light polarization

- *Light polarization* is the electric field vector orientation of a light wave.



Two-photon polarization microscopy (2PPM)

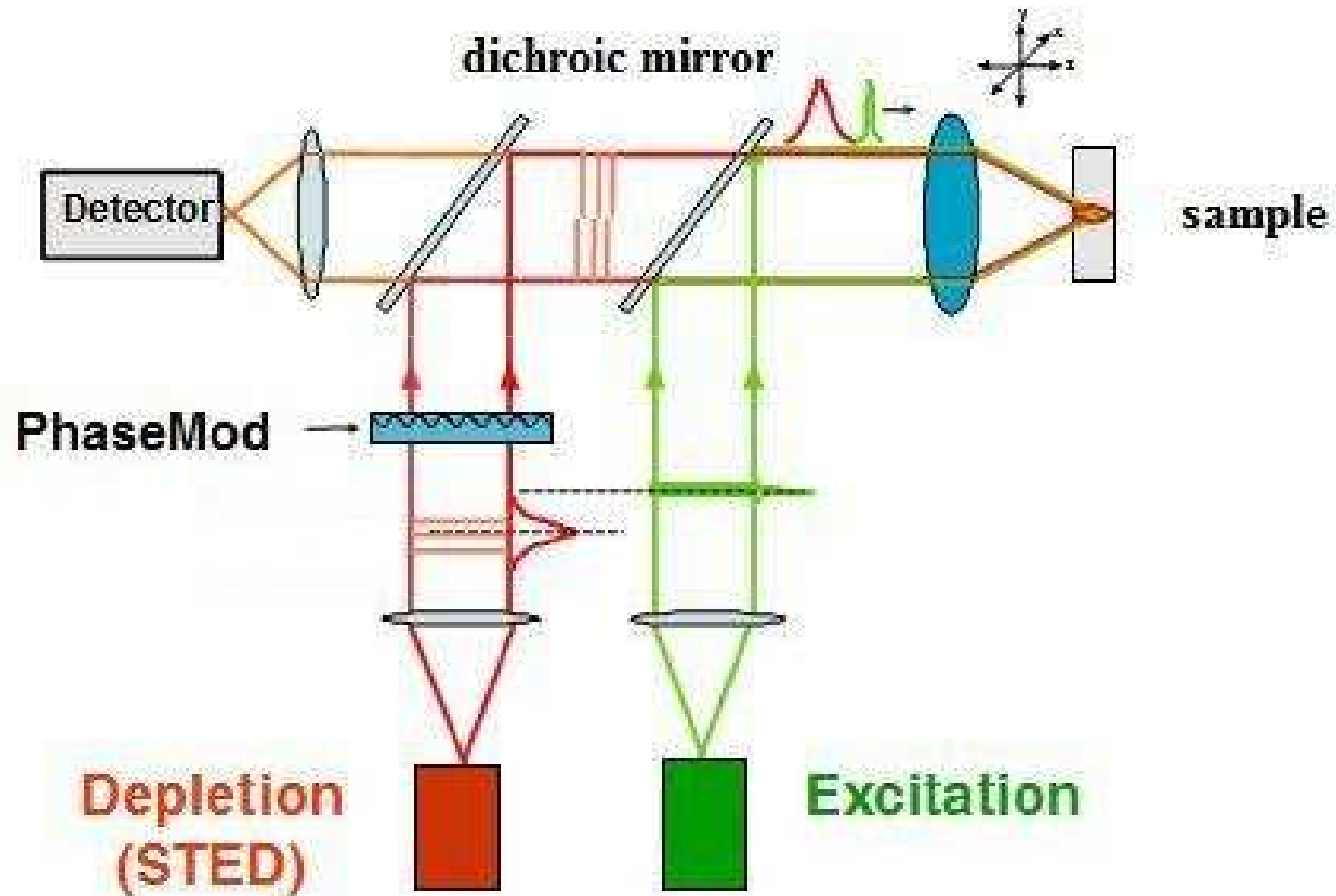
- A method of live cell microscopy which utilizes an optical property of fluorescent proteins called linear dichroism for determination of protein orientation with respect to the cell membrane.
- - live cell microscopy,
- - detects linear dichroism of fluorescent proteins,
- - works for membrane proteins (requires membrane),
- - shows protein orientation with respect to the membrane.

STED and RESOLFT

(stimulated emission depletion)

- STED technique utilizes two beams: excitation and doughnut-shaped depletion beam, to excite fluorescence in an area smaller than diffraction limit.
- RESOLFT technique uses photoswitchable fluorescent proteins in order to obtain superresolution.

STED setup



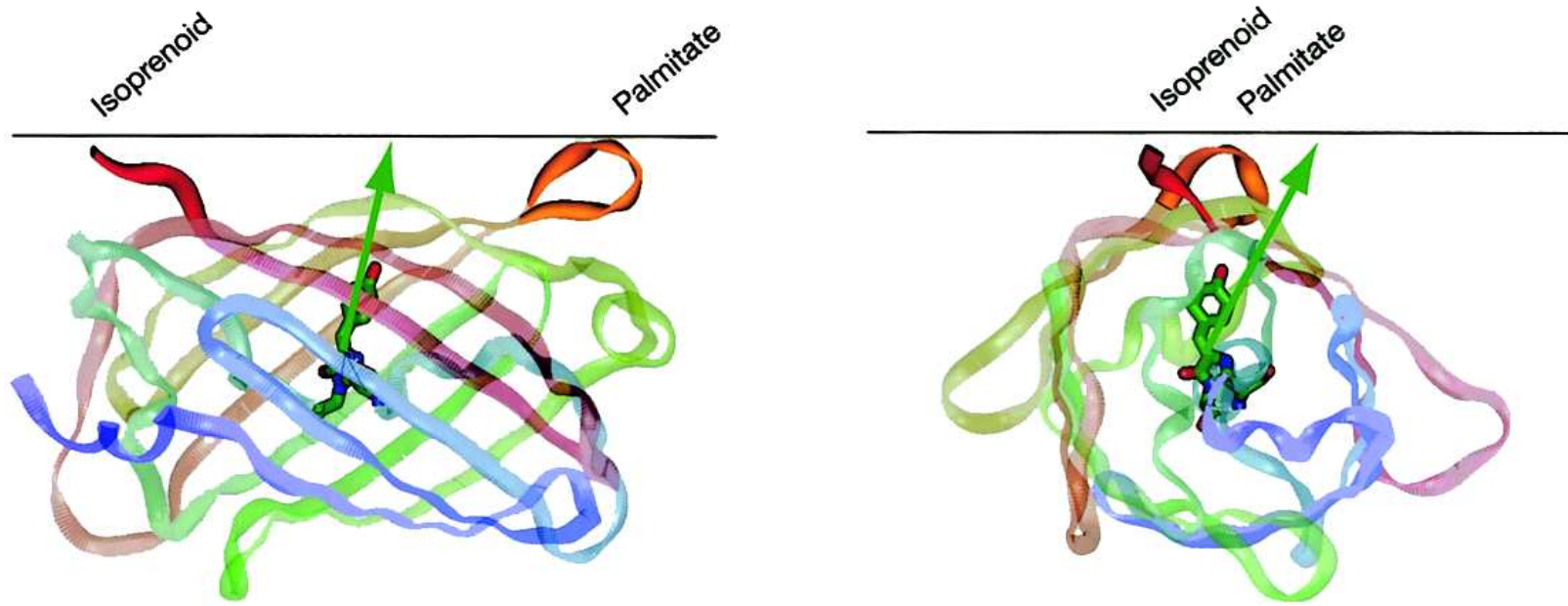
What is required for studies of the membranes?

- Need a protein which is:
- In the membrane or
- Is attached to the membrane.

- Original GFP- cytoplasmic localized.
- dleGFP.

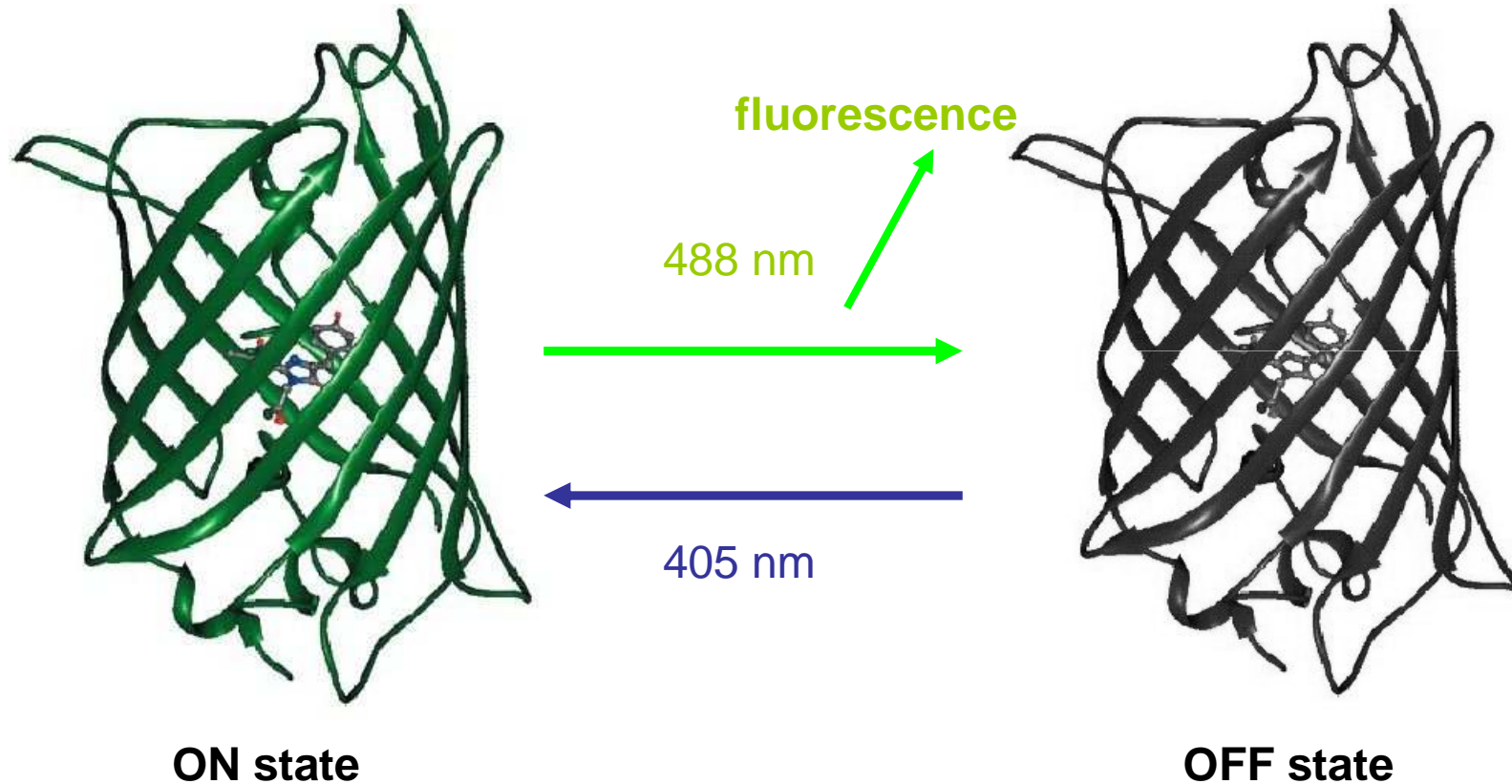
dIeGFP

- Doubly lipidated.
- Model construct for observation of eGFP linear dichroism in live cells.



(Roorda et al J. Neurophysiol. 2004)

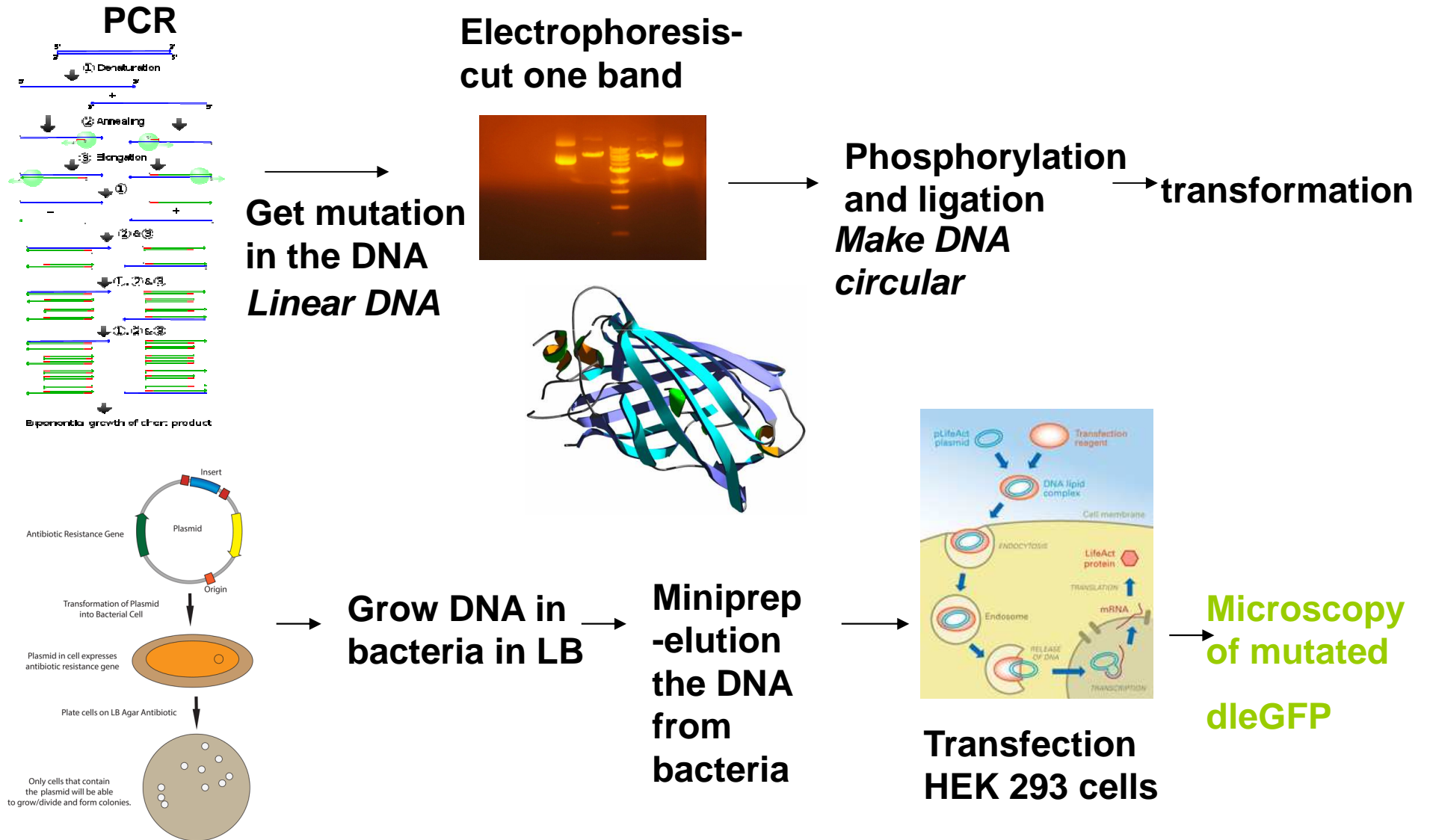
Photoswitchable GFP



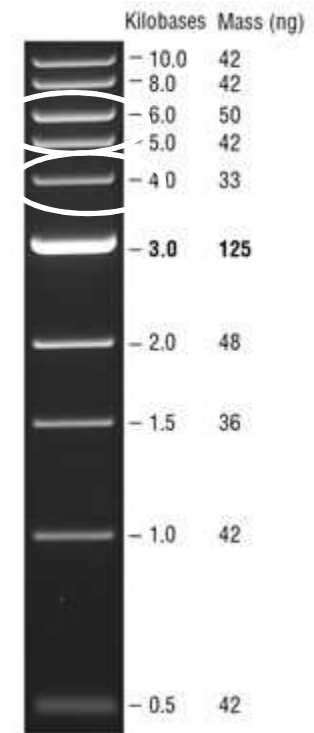
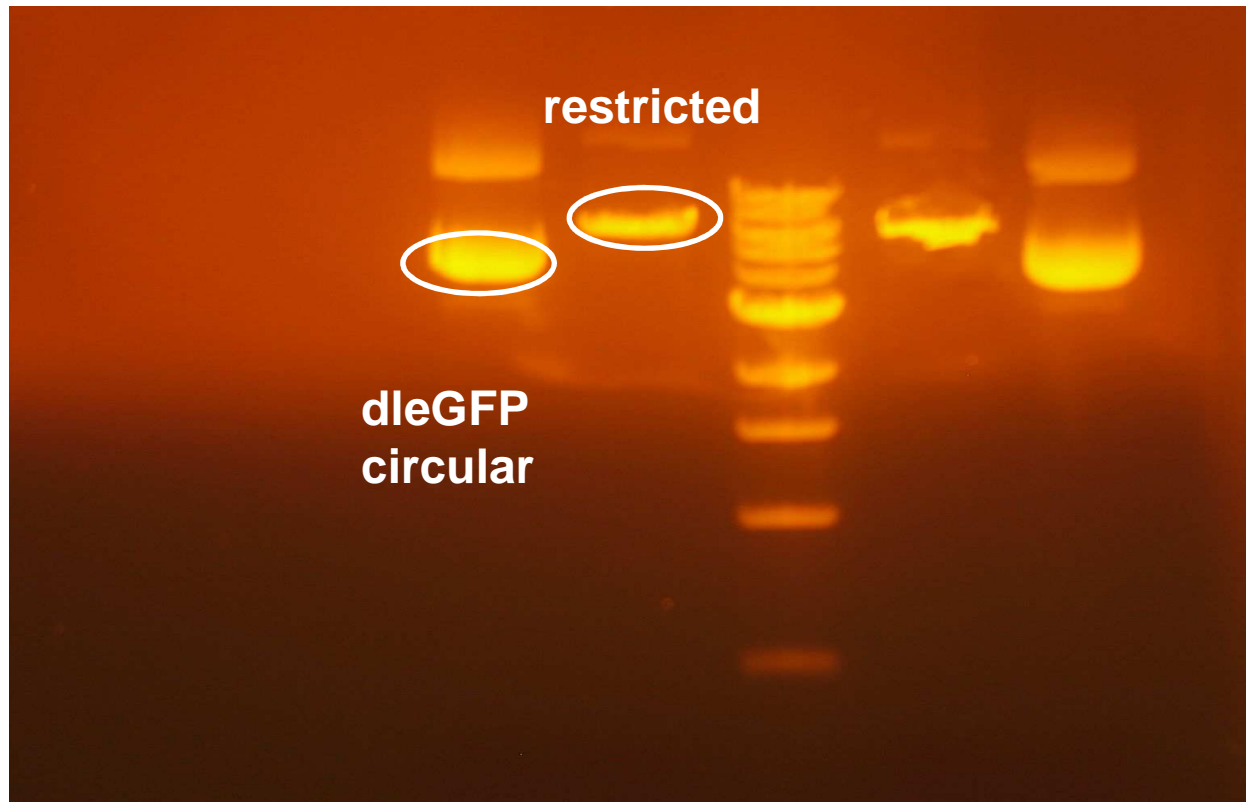
Experimental steps

- 1. Creating the mutant construct.
- 2. Testing the fluorescence properties.
- 3. Measuring linear dichroism of the mutant.

1. Creating the mutant construct



Original template and restriction with BaeI



Mutation of dleGFP

- Photoswitchable protein.
- EGFP[T65A, Q69L, V163S, A206K]
- PCR (Phusion Hot Start)

(Grotjohann, 2012)

PCR with primers A206K, T65A and Q69L

- A206K.

5'CTTATGGTGTTCTATGCTTTTCAA3(F),

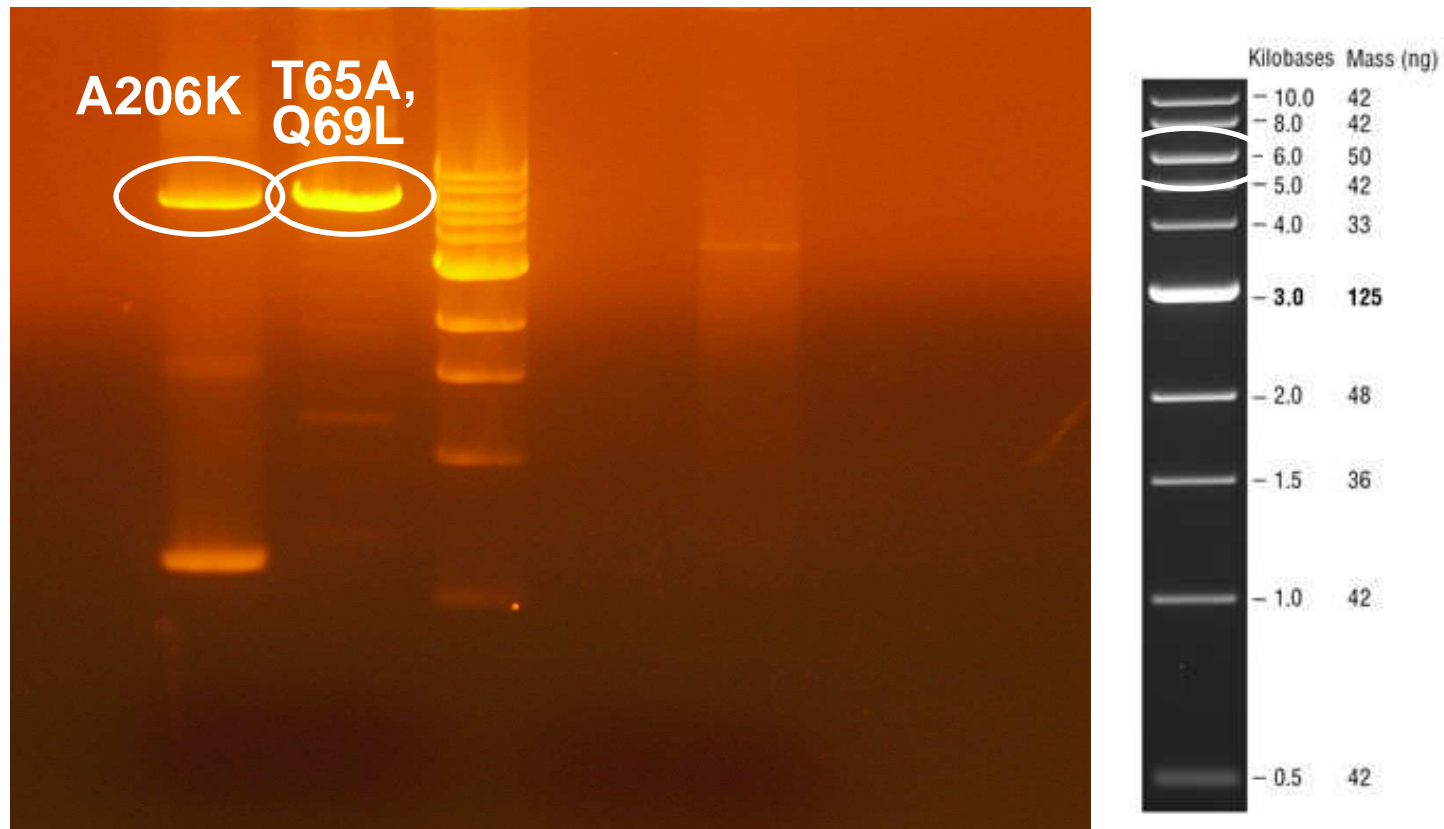
5'CTAAAGTAGTGACAAGTGTTGGC 3(R),

- T65A, Q69L.

5'CTTATGGTGTTCTATGCTTTTCAA3(F),

5'CTAAAGTAGTGACAAGTGTTGGC3,(R).

Mutation A206 K, T65A and Q69L



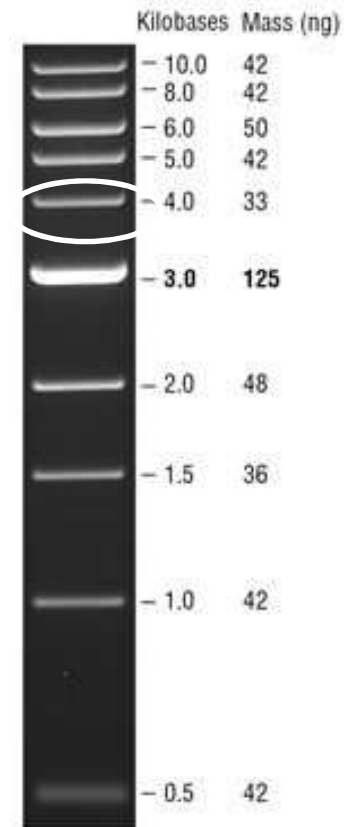
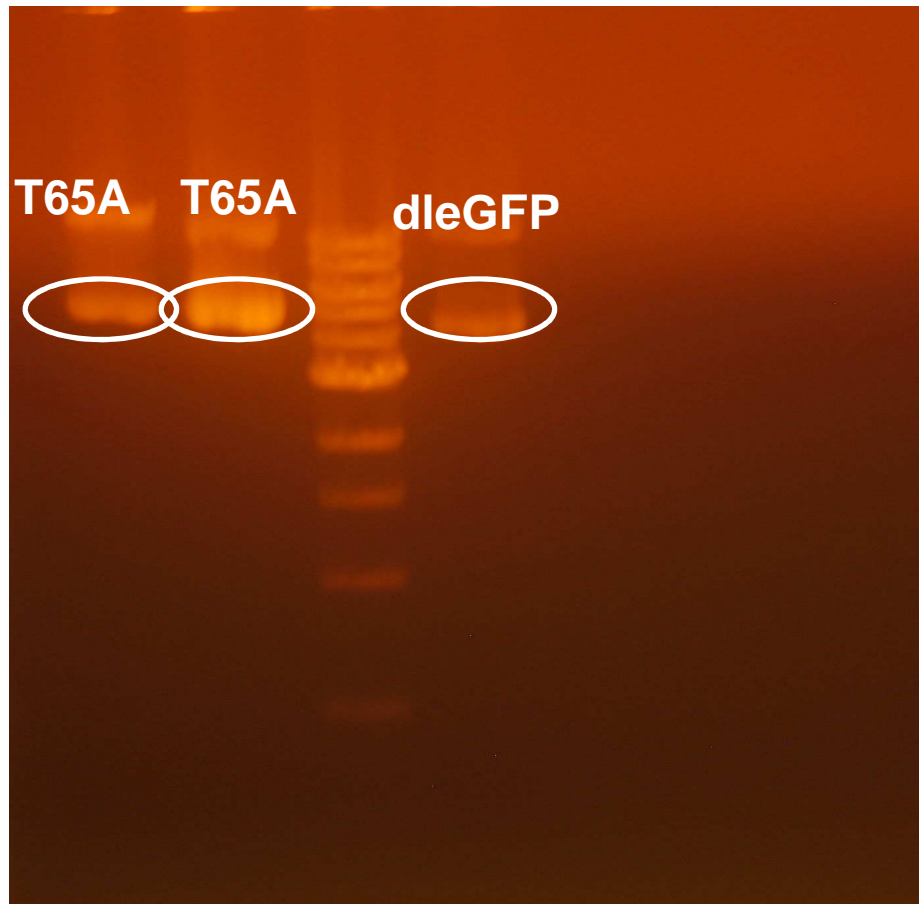
Gel extraction

- Cut one band from gel,
- Extraction from gel (extraction kit),



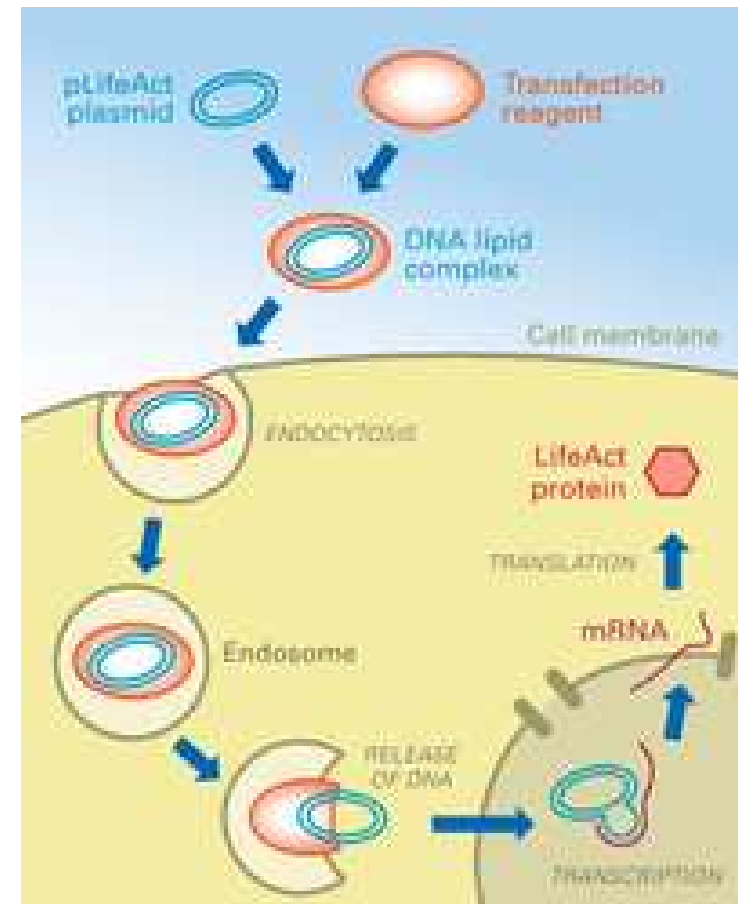
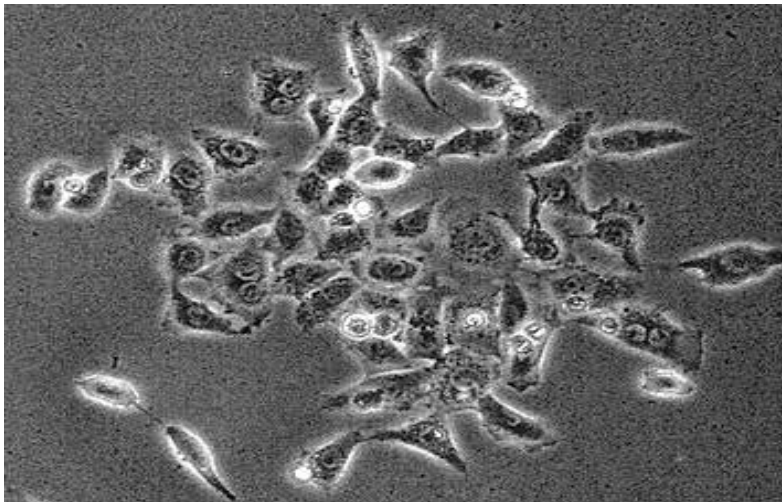
<http://www.qiagen.com/Products/Catalog/Sample-Technologies/DNA-Sample-Technologies/DNA-Cleanup/QIAquick-Gel-Extraction-Kit>

Electrophoresis after miniprep



Transfection in mammalian cells

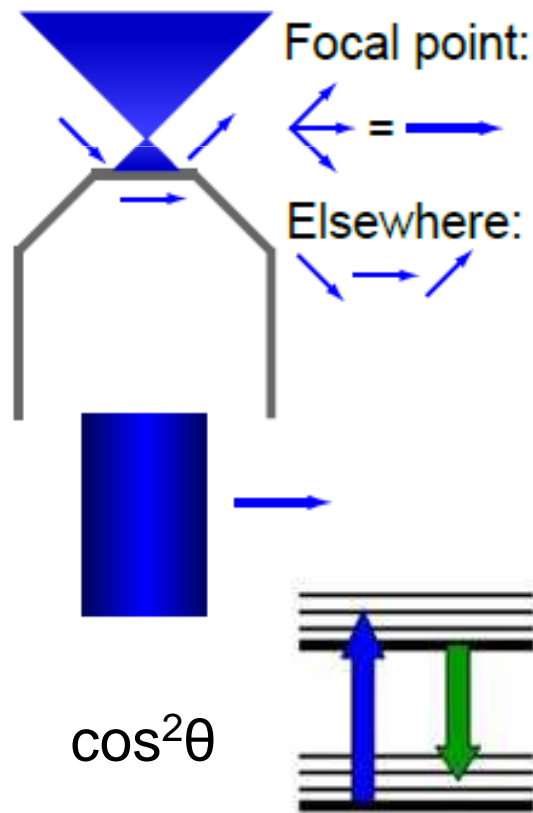
plasmid-> liposome->
endocytosis->core->
transcription->
translation->protein



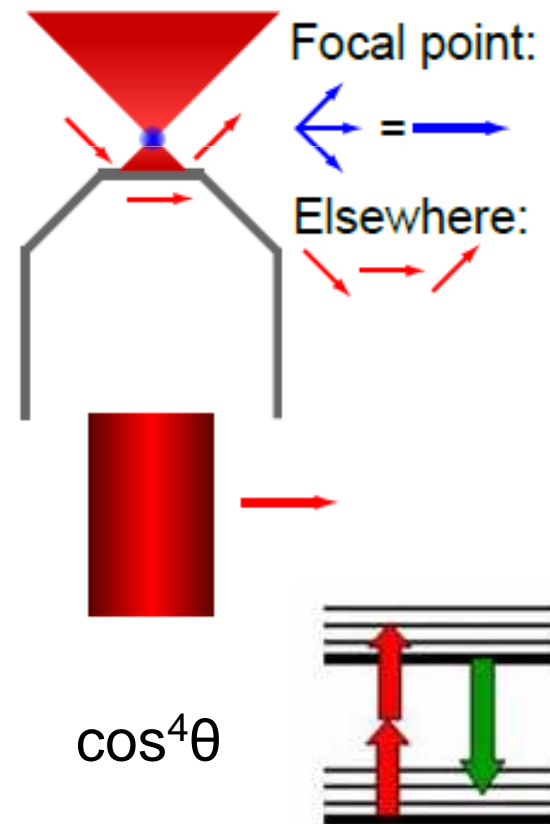
<http://ibidi.com/applications/transfection-transduction-and-proteofection/transfection/>,
http://en.wikipedia.org/wiki/File:HEK_293_cells_grown_in_tissue_culture_medium.jpg

Comparison of one-photon and two-photon microscopy

One-photon excitation



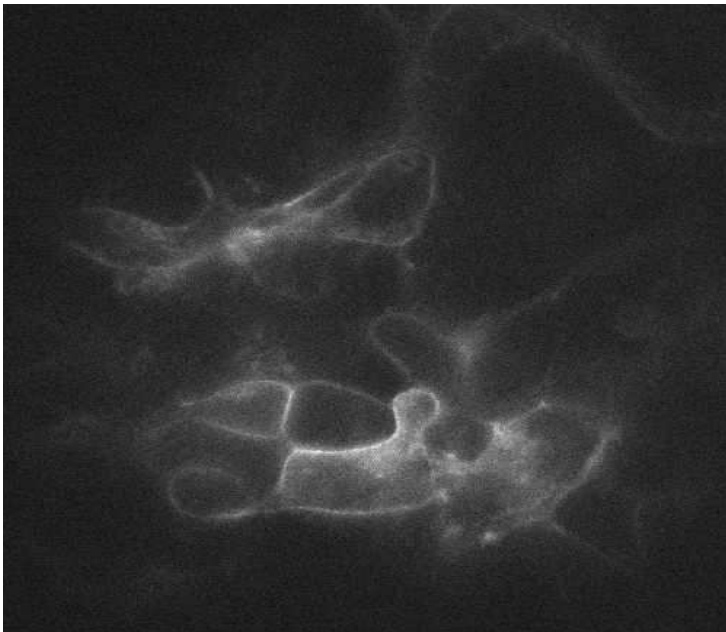
Two-photon excitation



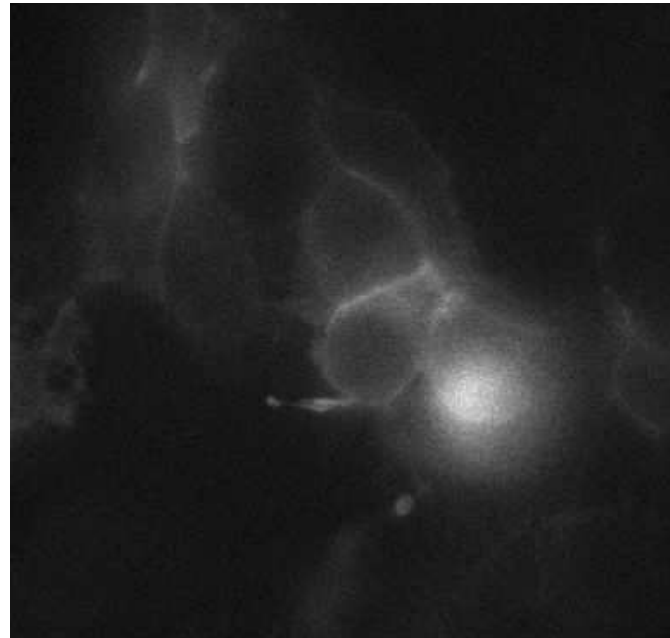
One photon microscopy

- Two wavelengths:
- 405 nm and 488 nm
- Zoom 60x.

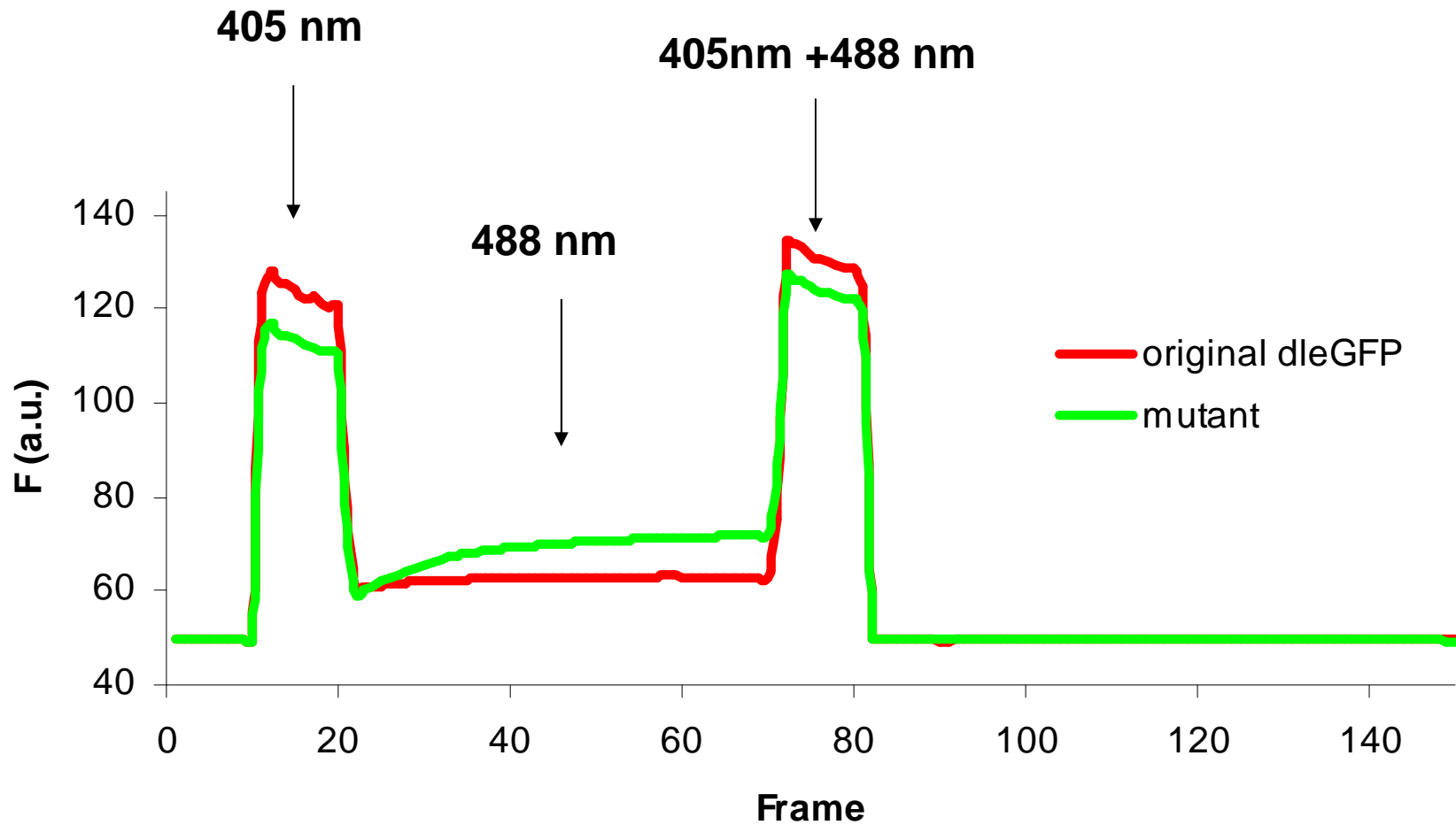
Original dleGFP



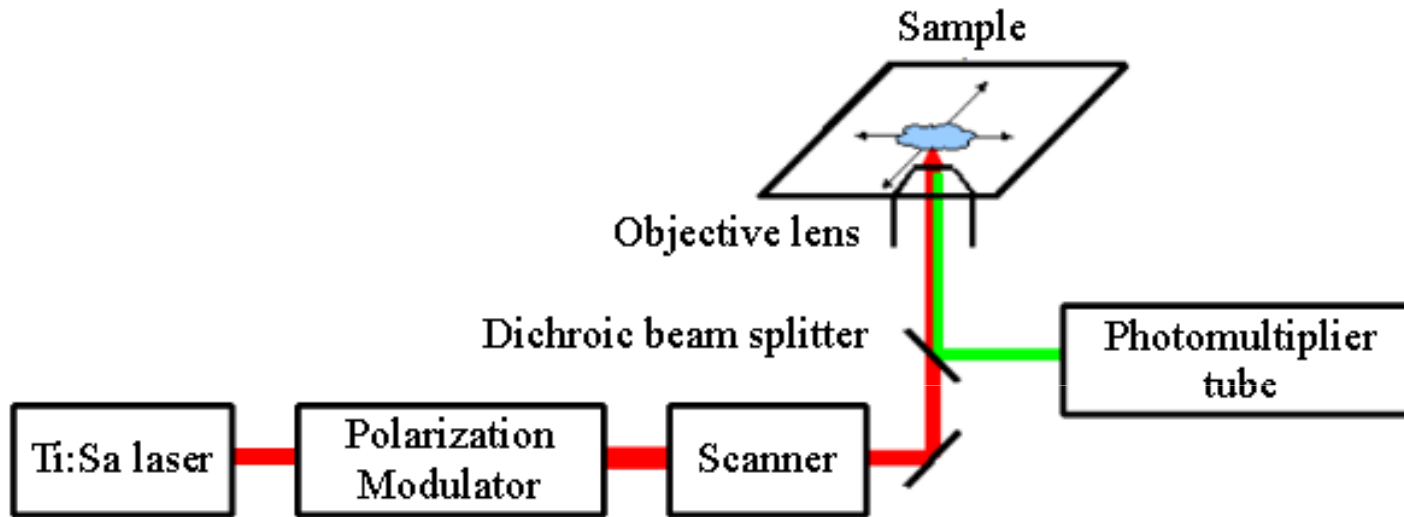
mutated dleGFP



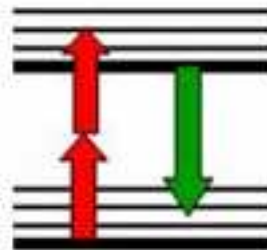
2. Testing the fluorescent properties



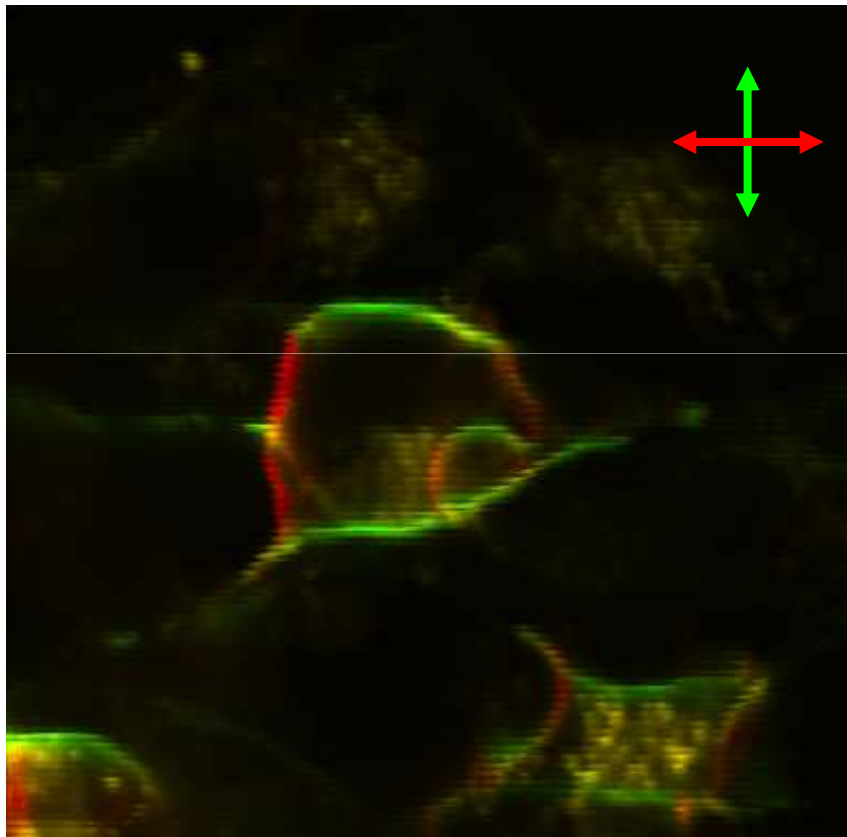
3. Two-photon polarization microscopy setup



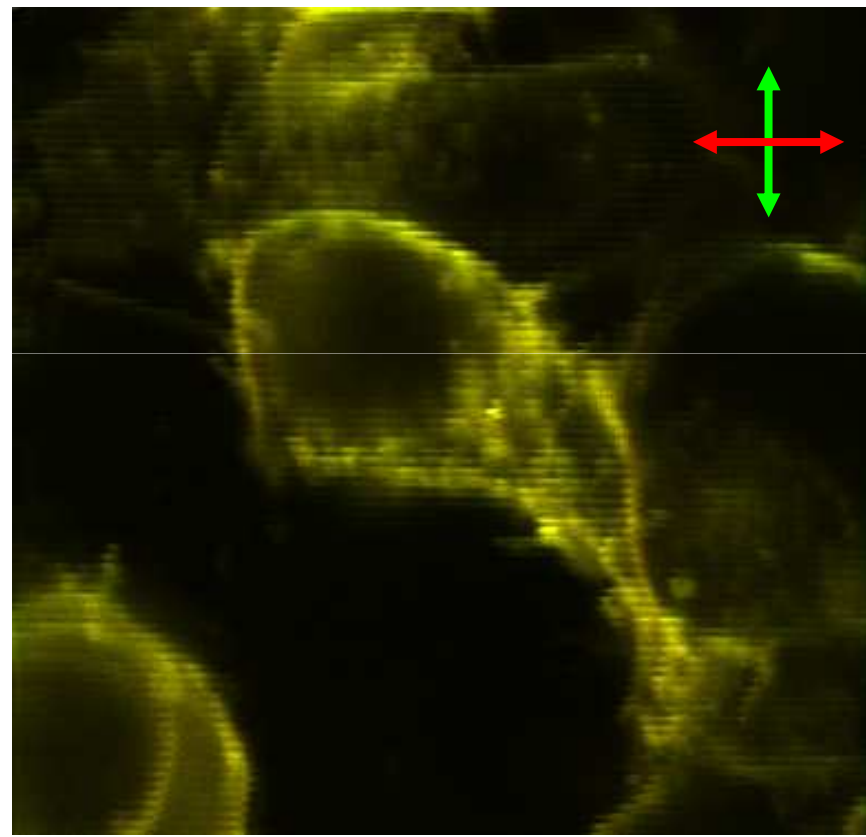
Two-photon excitation



2PPM of dleGFP and our mutant



dleGFP



T65A_G63A_dleGFP

Conclusions

- We introduced two point mutations (T65A and Q69L) in the dleGFP.
- Introduced mutations altered fluorescent properties of dleGFP and led to reduction of its linear dichroism due to changes of dleGFP fluorophore.
- Further work is required for introduction of two more mutations for improvement of photoswitching characteristics of dleGFP.

Acknowledgment

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